



# ***Pulse Quality Program Testing Manual***

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Ongoing - Version 3 - 2024

FOOD CROP QUALITY PROGRAM

SASKATCHEWAN FOOD INDUSTRY DEVELOPMENT CENTRE INC.



## **ACKNOWLEDGEMENTS**

We would like to express our sincere thanks to the Saskatchewan Pulse Growers for financially supporting this program.

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## **Appendix A: Pulse Quality Testing Manual**

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## 1. INTRODUCTION

The Pulse Quality Program launched in spring 2022 with a partnership between Saskatchewan Pulse Growers and the Saskatchewan Food Industry Development Centre, Inc (Food Centre). The mission of the program is to add in best management practices for pulses grown in Western Canada and to help the development of pulse-based ingredients/products in the food industry. The program will develop a comprehensive database of composition, functionality, and nutrition for pulses and related standardized methodology for high throughput screening purposes.

Pulses are being recognized as excellent food sources to provide high proteins, dietary fibers, and a sufficient amount of minerals and thereby have gained stupendous interests in the supply chain (Tosh & Yada, 2010)<sup>1</sup>. In the past decade, pulse-related products have increased by over five times, and pulse flours have been mainly applied in the extruded (pasta, noodles, snacks), baked (bread, cakes, cookies, gluten-free products), and meat-related products in the food industry. Canada is one of the largest producers of pulses and the global leader in pulse exports. The production of national peas and lentils in 2020 was 4.6 and 2.9 million tonnes, respectively, where Saskatchewan has contributed 84-96% and 48-63% of lentil and pea production over the past decade (Canadian Grain Commission, 2011-2021).

A genotype by environment evaluation (GxE) on quality parameters of pulses will give information to growers, breeders, pulse processors, and end users. For instance, information can help growers and breeders monitor their crops, change their decision making on cultivation of pulse crops the next year, and lead to new breeding program. Knowledge will help pulse processors on ingredient selection and product development. Data will also help SK Food Centre develop value proposals for the pulse processing industry

This program will implement a GxE evaluation on pulse quality. In the first phase of the program, pulses including peas, lentils, chickpeas, faba beans, and dry beans with up to 3000 samples annually from multiple cultivars and locations in 2021-2024 will be analyzed. The main focus of parameters includes seed quality (i.e. 1000 seed weight, amount of damage and foreign matter, seed size and hardness, and appearance), nutritional composition (i.e. ash, moisture, and protein contents), and physical properties (i.e. colour, particle size, and Hausner ratio). This testing manual describes the procedure to evaluate each parameter above.

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<sup>1</sup> Tosh, S. M., & Yada, S. (2010). Dietary fibres in pulse seeds and fractions: Characterization, functional attributes, and applications. *Food research international*, 43(2), 450-460.

## **2. TESTING PROTOCOLS**

### **A. Sample labelling**

Sample labelling is critical when handling a significant number of samples. In this project, samples from the same type, year, and location are determined at a time for all the experiments. Thus, samples to be measured at the same time is labelled according to the plot number.

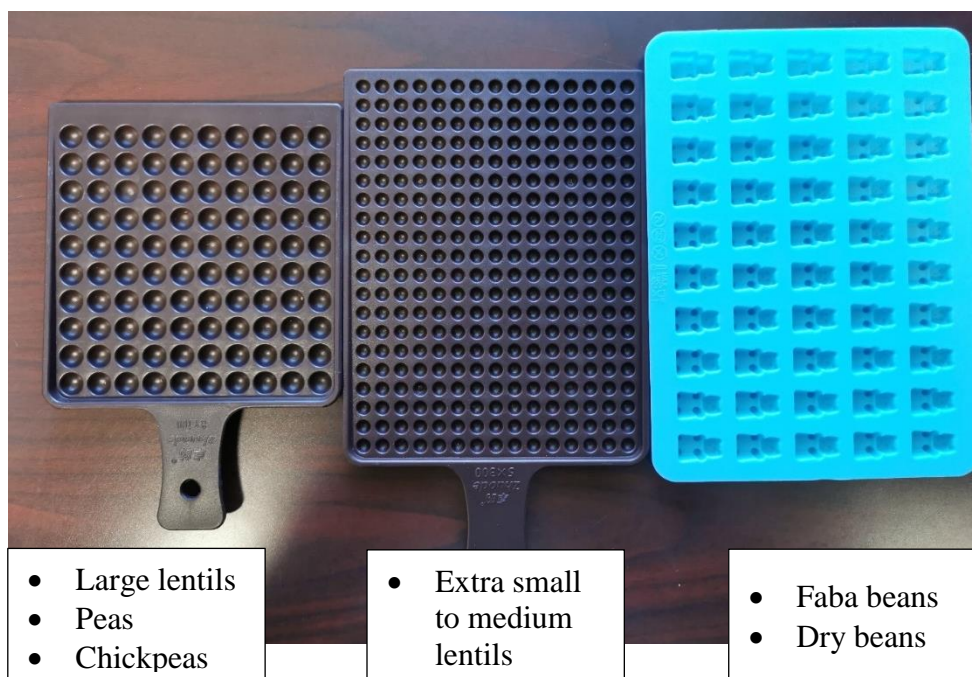
Results are recorded in the template Excel sheets, which can be found in the file of “Food Crop Quality Program” under Group Storage “G” drive.

## B. 1000 Seed weight

Seed weight is an important parameter to indicate seed size and yield production. Knowledge of this characteristic is fundamental to seeding management and pulse crop establishment. This method determines the weight of 1000 seeds.

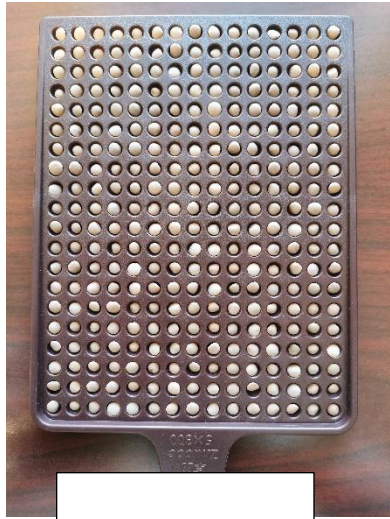
### Apparatus:

1. Top loading balance
2. Counting trays



### Procedure:

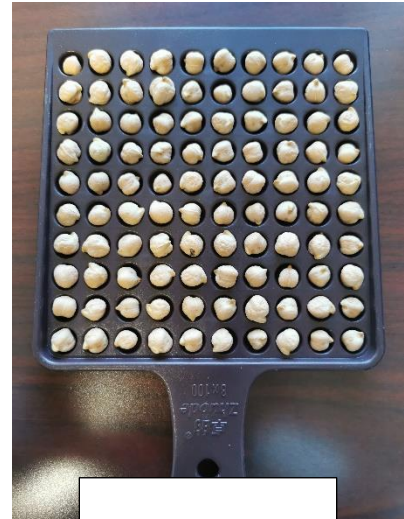
1. Place one seed to each mold of the counting trays until all the molds are filled with seeds.
  - a. **300/ pan seed counting tray:** Lentil (extra small, small, French green).
  - b. **100/ pan seed counting tray:** Lentil (medium, large); Pea; Chickpea.
  - c. **Gummy bear seed counting tray:** Faba bean; Dry bean.
2. Repeat step 1 to achieve 300 seeds. Weigh 300 seeds and record. Broken and damaged seeds are excluded.
3. Duplicated measurements are preformed. The 1000 seed weight is calculated and reported using the equation below.



- Small lentils



- Green peas



- Chickpeas

**Calculation:**

$$1000 \text{ seed weight (g)} = \frac{300 \text{ seed weight (g)}}{3} \times 10$$



### C. Seed size distribution

This method determines the size distribution of the pulse seeds. Seed size is important in seeding management and pulse crop establishment.

#### Apparatus:

1. Metal hand sieves
  - a. #28R: 11.11 mm
  - b. #26R: 10.32 mm
  - c. #24R: 9.52 mm
  - d. #22R: 8.73 mm
  - e. #20R: 7.94 mm
  - f. #18R: 7.14 mm
  - g. #16R: 6.35 mm
  - h. #14R: 5.56 mm
  - i. #12R: 4.76 mm
  - j. #10R: 3.97 mm



#### Procedure:

1. Weigh  $250 \pm 2$  g seeds and record. Working sample size and sieve sizes are chosen according to the recommendation of Official Grain Grading Guide.
2. Pile the sieves from the largest opening to the smallest opening.
3. Pour the sample seeds from the top sieve, and shake until all the seeds pass through the openings of each sieve.
4. Take the top sieve to a second container. Shake again and make sure the smaller seeds pass through the sieve. Pour the retained seeds of each sieve into a separate container and pick out the foreign matter.
5. For picking the foreign matter, collect all seeds into a deli cup and then pour a very small amount at a time into a larger container. Pick out the foreign into a separate deli cup and then swipe the lentils to the side in the container. Repeat until all lentils are cleaned of foreign.
6. Weight the retained seeds.
7. Repeat steps 4 to 6 for the next sieve.
8. Collect all the foreign matter at the end and record the total weight.
9. The above measurements are to be duplicated for the same cultivar and same location.

#### Sieve selection:

**Peas:** green, maple, and yellow.

- Green: normally size from #20R (7.94 mm) to below #14R (5.56 mm)
- Yellow: normally size from #20R (7.94 mm) to below #14R (5.56 mm)

- Maple: normally size from #20R (7.94 mm) to below #14R (5.56 mm)
- If over 10 g of peas remain at #20R (7.94 mm) sieve, please sieve through using the #22R (8.73 mm) sieve as well.
- Anything passing through #14R (5.56 mm) is considered smaller.

**Lentils:** Green (French green, small, large), Red (extra small, small, medium, large), and Spanish brown.

- Extra-small: normally size from #14R (5.56 mm) to below #10R (3.97 mm)
- Small: normally size from #14R (5.56 mm) to below #10R (3.97 mm)
- Medium: normally size from #16R (6.35 mm) to below #10R (3.97 mm)
- Large: normally size from #18R (7.14 mm) to below #10R (3.97 mm)
- If over 10 g of lentils remain at the top sieve, please try it with a larger sieve.
- Anything passing through #10R (3.97 mm) is considered smaller.

**Chickpeas:** black desi, desi, and kabuli.

- Black desi: black color, normally size from #20R (7.94 mm) to below #14R (5.56 mm)
- Desi: brown color, normally size from #22R (8.73 mm) to below #14R (5.56 mm)
- Kabuli: yellow color, normally size from #24R (9.52 mm) to below #14R (5.56 mm)
- If over 10 g of seeds remain at the top sieve, please try it with a larger sieve.
- Anything passing through #14R (5.56 mm) is considered smaller.

**Faba beans:** Tannin (coloured flower) and Zero tannin (white flower)

- Tannin: normally size from #28R (11.11 mm) to below #14R (5.56 mm)
- Zero tannin: normally size from #26R (10.32 mm) to below #14R (5.56 mm)
- If over 10 g of seeds remain at the top sieve, please try it with a larger sieve.
- Anything passing through #14R (5.56 mm) is considered smaller.

**Beans:** Black, FDJ, navy, pinto and yellow

- Navy: normally size from #14R (5.56 mm) to below #10R (3.97 mm)
- Black: normally size from #16R (6.35 mm) to below #10R (3.97 mm)
- FDJ, pinto and yellow: normally size from #22R (8.73 mm) to below #10R (3.97 mm)
- If over 10 g of beans remain at the top sieve, please try it with a larger sieve.
- Anything passing through #10R (3.97 mm) is considered smaller.

**Calculation:**

For instance, for seeds do not pass through #24 (9.52 mm):

$$>9.52 \text{ mm } (\%) = \frac{\text{weight of seed collected at \#24 (g)}}{\text{sum of seed weight (g)}} \times 100\%$$

## D. Amount of foreign matter

The presence of foreign matter reduces pulse value. Information of this characteristic will help determine the pre-harvest factors and harvest method for pulse harvesting. This protocol determines the purity of the seeds.

### Apparatus:

1. Top loading balance

### Procedure:

1. Foreign matter is selected out during the measurements of sizing distribution.
2. Pick out the foreign matter remained at each metal hand sieve.
3. Weigh out the total foreign matter. Duplicated measurements are performed, and the average is reported.

### Calculation:

$$\text{Foreign matter (\%)} = \frac{\text{weight of foreign matter (g)}}{\text{sum of seed weight and foreign matter (g)}} \times 100\%$$

### Note:

1. Foreign matter includes **ergot, excreta, insect parts, stones, other type or cultivar of seeds, and all other extraneous materials.**
2. Pea:
  - a. When sizing green peas, please differentiate between bleached green pea (appears as yellow) and yellow pea. When sizing yellow peas, please differentiate between immature yellow pea (appears as green) and green pea.
3. Lentil:
  - a. A lentil is prone to contain other types of lentils in the plot due to contamination from harvesting.
  - b. Key features to differentiate: **speckles, seed coat colour, and size.**
  - c. A similar colour seed coat/hull makes different types of lentils difficult to identify from each other. Please verify with supervisor when in doubt.
  - d. If you are unsure whether a lentil is foreign, the lentil can be crushed with mortar and pestle or snipped with scissors a small amount of the outer seed coat to see the cotyledon colour.
4. Chickpea, faba bean, and dry bean:
  - a. Key features to differentiate: **seed coat color and size.**

## E. Amount of damage and splits

The presence of damaged seeds reduces pulse value. Information on this characteristic will help determine the pre-harvest factors and harvest method for pulse harvesting. This protocol determines the damage of the seeds.

### Apparatus:

1. Top loading balance

### Procedure:

1. Weigh out the recommended seed weight (see Table 1e) and record the weight. Working sample size is chosen according to the recommendation of Official Grain Grading Guide<sup>2</sup>.
2. Select out the damaged seeds and splits by hand, and weigh them separately.

**Table 1e.** Working sample size of each seed type for damage and split determination.

Seed type	Working sample size (g)
Lentil	50
Pea	100
Chickpea	100
Faba bean	100
Dry bean	100

### Calculation:

$$\text{Total damage (\%)} = \frac{\text{weight of damaged seed (g)}}{\text{weight of seed (g)}} \times 100\%$$

$$\text{Split or mechanical damage (\%)} = \frac{\text{weight of splits (g)}}{\text{weight of seed (g)}} \times 100\%$$

$$\text{Other damage (\%)} = \frac{\text{all types of damage excluding mechanical damage (g)}}{\text{weight of seed (g)}}$$

**Mechanical damages** are (but not limited to): splits, cracks, partially missing hull, and partially missing cotyledon.

- A deformed seed is NOT considered as damage.

<sup>2</sup> Canadian Grain Commission. (2023). <https://www.grainscanada.gc.ca/en/grain-quality/official-grain-grading-guide/> Access date: Jan 03, 2024



- Seed coat or cotyledon loss with less than 20% of the surface area is NOT considered as damage.

### Other damage:

#### Pea:

Other damage are (but not limited to): sprouted, shrivelled, heated, and insect damaged.

- **Shrivelled:** shrunken or dimpled surface.
- **Heat-damaged:** dull seed coats and discoloured cotyledons from light tan to dark brown.
- **Insect-damaged:** damaged by insects.
- **Sprouted:** when the seed oat splits and the primary sprout emerges from between the cotyledons.
- **Pink peas:** caused by bacteria *Erwinia Rhapontici*. Be able to differentiate pink pea (disease) and red pea (contamination).
- **Bleaching:** natural sunlight damage that only applies to green peas. When 1/8 or more of the surface of the cotyledon is bleached to a distinct yellow colour.
- **Wrinkles:** caused by rapid drying from processing during harvesting.
- **Red peas (contamination):** pay attention to red/orange peas for varieties CDC 5779-1, CDC 5791-9, and CDC 5845-2 from 2022



**Figure 1e.** Example of wrinkled damage, water damage, red (contamination), and pink (disease) peas (from left to right).

#### Lentils:

Other damage are (but not limited to): sprouted, frost damaged, heated, insect damaged, wrinkled, distinctly green, and disease.

- **Wrinkled:** ONLY apply to red lentils. Recognize as wrinkles if the surface has a sharp ridge. Consider as sound for a dimpled seed coat.
- **Sprouted:** when the seed oat splits and the primary sprout emerges from between the cotyledons.
- **Heat-damaged:** appears as dark tan to black.
- **Insect-damaged:** damaged by insects.

- **Frost-damaged:** indicated by a combination of wrinkling and close adherence of the seed coat to the cotyledon.
- **Disease/Ergot:** a purplish-black exterior, a purplish-white to off white interior, and a relatively smooth surface.

### **Chickpeas:**

Other damage are (but not limited to): sprouted, frost damaged, heated, mouldy, un-removable dirt, and insect damaged.

- **Dirt:** try to remove it with your fingers. Consider it as damage if the dirt is not removable and over 50% of the surface area.
- **Sprouted:** when the seed coat splits and the primary sprout emerges from between the cotyledons.
- **Heat-damaged:** dull seed coats and discoloured cotyledons from light tan to dark brown.
- **Frost-damaged:** green or white and shriveled.
- **Green:** if **Kabuli type** shows any green colour on the seeds or seed coats and if **Desi type** shows distinctly green colour on the cotyledons.
- **Mouldy:** if they show clear evidence of mildew or mould.

### **Faba beans:**

Other damage are (but not limited to): sprouted, frost damaged, heated, insect damaged, wrinkled, distinctly green, and disease.

- **Sprouted:** when the seed coat splits and the primary sprout emerges from between the cotyledons.
- **Heat-damaged:** appears as dark tan to black.
- **Insect-damaged:** damaged by insects.
- **Frost-damaged:** indicated by a combination of wrinkling and close adherence of the seed coat to the cotyledon.
- **Disease/Ergot:** a purplish-black exterior, a purplish-white to off white interior, and a relatively smooth surface.
- **Blackened:** when their seed coats are very dark blue to black.
- **Distinct immaturity:**
- **Mould damage:** if they show clear evidence of mildew or mould.



**Figure 2e:** Damage caused by lygus bugs.



**Figure 3e:** Sprouting of Navi faba beans from water imbibition.



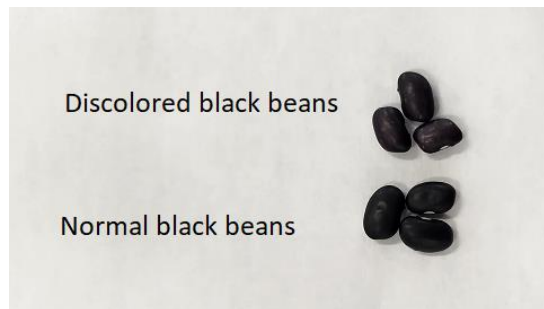
**Figure 4e:** Sprouting of DL Nevado.

### **Dry beans:**

Other damage are (but not limited to): sprouted, frost damaged, heated, insect damaged, mouldy, rotted, and disease.

- **Heat-damaged:** appears as dark tan to black.
- **Insect-damaged:** damaged by insects.
- **Disease/Ergot:** a plant disease producing elongated fungus bodies with a purplish-black exterior, a purplish-white to off white interior, and a relatively smooth surface.

- **Mouldy:** presence of dark blue exterior moulds that develop in machine-damaged crevices. For instance, light and dark red kidney beans may develop yellow to black interior moulds in the concave centre area.
- **Rotted:** whole beans or pieces of beans that are visibly in advanced stages of decomposition and that feel spongy under pressure.
- **Discolouration:** when discolouration on the seed coat covers more than half the bean or when the discolouration penetrates the cotyledon.



**Figure 5e.** Discoloration of black bean.



## F. Seed hardness

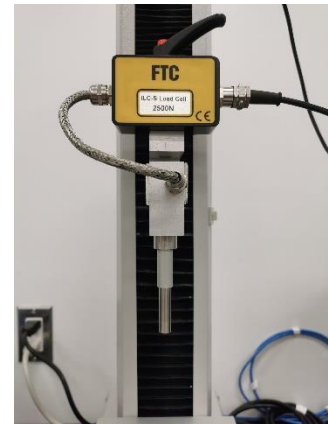
Seed hardness is an important parameter to indicate milling yield and cooking quality. This method determines the hardness of pulse seeds by measuring the crushing strength of individual seeds using a texture analyzer (TMS-Pro, Food Technology Corporation, USA) equipped with a 2500 N load cell with a modified method from Karami et al. (2017) and Lovas-Kiss (2020)<sup>3</sup>.

### Apparatus:

1. Texture analyzer (1 cm probe and 2500 N loading cell)
2. Cleaning brushes

### Procedure:

1. Make sure the loading cell is **2500 N** and put the **1 cm probe** to the loading cell.



2. Turn on the texture analyzer.
3. Click on software **Texture lab Pro** (name & password: admin).
4. Click **ok** for program testing.
5. In menu, click on **file > load library program > seed hardness-(seed type)**. Click open.
6. Click **zero load** to zero force.

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<sup>3</sup> Karami, S., Sabzalian, M. R., Rahimmalek, M., Saeidi, G., & Ghasemi, S. (2017). Interaction of seed coat color and seed hardness: An effective relationship which can be exploited to enhance resistance to the safflower fly (*Acanthiophilus helianthi*) in *Carthamus* spp. *Crop Protection*, 98, 267-275.

Lovas - Kiss, Á., Vincze, O., Kleyheeg, E., Sramkó, G., Laczkó, L., Fekete, R., ... & Green, A. J. (2020). Seed mass, hardness, and phylogeny explain the potential for endozoochory by granivorous waterbirds. *Ecology and Evolution*, 10(3), 1413-1424.

7. Move the probe to the zero position (i.e., move it as close as possible to the platform with force remaining at 0.0 N). Click **zero displacement**. You should try a few times to make sure the zero position is correct.
8. Move the probe to the recommended starting position (Table 1f); then click **zero displacement** again.
9. Click on **test program**. Make sure the program setting is the same as Table 1f.
10. Click on **setup > preference > general**. Make sure **compression** is chosen and **2500 N** is set as defaulted.
11. Place the seed right under the probe. Make sure **the seed is at the center** by checking from at least two positions.
12. Click **start** and record the peak value.
13. Report the average of 10 measurements for each sample.
14. Save the file to: G drive>Food Crop Quality>Seed-Year>Location-Seed-Year>Texture Raw Data

**Note:**

1. The probe can be moved to higher than 10 mm for a new starting position depending on the seed size in step 8.

**Table 1f.** Program setting for each type of seed.

Seed type	Starting position Distance in step 6 (mm)	Break point	
		Distance (mm)	Break percentage (%)
Lentil	8	1.5	30
Pea	10	7	50
Chickpea	10	7	25
Faba bean	10	7	50
Dry bean	10	6	50

## G. Appearance of whole seed

This procedure describes the steps to take a photo of each pulse seed.

### Apparatus:

1. Photo taking case
2. Petri dish

### Procedure:

1. Fill up the pulse seed in the petri dish.
2. Place the dish in the centre of the photo case and turn on the light.
3. One representative photo for each sample is reported.

### Lentils



**Figure 1g:** Red lentils of small (left), medium (middle), and large (right).



**Figure 2g:** Green lentil of small (left), large (medium), and French green (right).



**Figure 3g:** Spanish brown lentils (note the speckles).

#### **Faba beans**



**Figure 4g:** zero tannin faba beans (left) and tannin faba beans (right).





**Figure 5g:** zero tannin faba beans: DL Nevado (left) and 1089-1-2 (right).

### Dry Beans



**Figure 6g:** Types of dry beans.

## H. Milling of flour

Milling of pulses refers to a combination of dehulling and flour milling (Thakur et al., 2019)<sup>4</sup>. Pulse quality attributes, such as moisture content and structure (i.e. hardness, size, and hull content), significantly impact the milling quality. Dehulling, which is the process of removing seed coats, increases the appearance, texture, palatability, and digestibility of seeds and removes 80-90% of anti-nutritional factors (Narasimha et al., 2003). Dehulling efficiency is an imperative parameter for breeders to monitor the pulse crop quality and for the pulse processors in selecting ingredients and developing new products. Flour milling indicates particle size reduction, so as to facilitate ingredient miscibility. It is critical for pulse to be milled into consistent, high-quality ingredients for the food industry. Milling all samples to the same level of fineness will help compare the flour quality and functionality.

### Note:

1. Make sure a mill is cleaned properly before starting the next milling.
2. Pay attention to the temperature of the mill. Stop milling and wait for 15 min when it gets too hot.
3. Make sure the milling yield is higher than 90%.
4. Wear the N95 mask and ear muff during milling process.

### Calculations:

$$\text{Milling yield (\%)} = \frac{\text{weight of flours (g)}}{\text{weight of seeds}} \times 100\%$$

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<sup>4</sup> Narasimha, H. V., Ramakrishnaiah, N., & Pratapa, V. M. (2003). Milling of pulses. In: Handbook of Post Harvest Technology. New York, USA.: Marcel Dekker, Inc.

Thakur, S., Scanlon, M., Tyler, R., Milani, A., & Paliwal, J. (2019). Pulse flour characteristics from a wheat flour miller's perspective: A comprehensive review. *Comprehensive Reviews in Food Science and Food Safety*, 18(3), 775-797.

**i. Pin mill**

Preliminary tests show that the pin mill is a good device for pre-breaking pulses, such as faba beans, chickpeas, dry beans, and peas.

**Apparatus:**

1. Pin mill
2. Balance
3. Brushes
4. Vacuum
5. Burr grinder
6. N95 mask
7. Ear plug or ear muff



**Procedure:**

1. Weigh out 210 g seeds (without foreign matter).
2. Close the chamber properly and plug the power.
3. Set the gap as the largest and turn on the mill. Pour about 40 g of samples from the top and wait for about 30 - 45 seconds. Listen the voice to determine whether milling is finished.
4. Unplug the power and collect all the coarse flours.
5. Open the mill and clean it using brushes and vacuum.

## ii. Cyclone mill

The Udy cyclone mill uses a high velocity air-flow, an abrasive surface, and centrifugal forces to grind material. The impeller rotates at a high speed creating the high velocity flow of air to propel articles against the abrasive surface. It is a good device to mill the pulses as a whole into fine powders. Milling yield of the Udy mill is 95%. The procedure describes the steps of milling using the cut mill.

### Apparatus:

1. Udy cyclone sample mill
2. Balance
3. Brushes
4. Vacuum
5. Burr grinder
6. N95 mask
7. Ear plug or ear muff

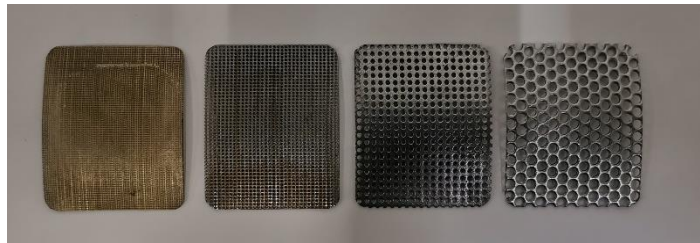
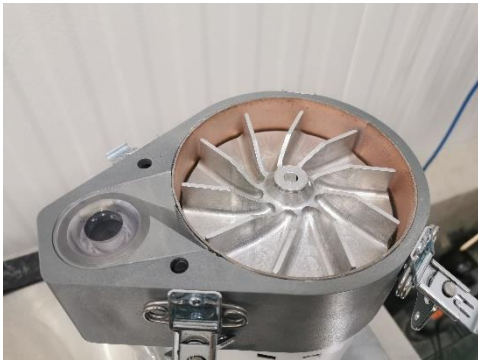


Figure 1. Udy mill screens: 0.25 mm, 0.5 mm, 1 mm, and 2mm (from left to right).

### Procedure:

1. Weight 210 g of whole seeds or coarse flours.
2. Verify that a screen, an impeller, and the cyclone air separator are in place.
3. Place the Cover on the top of the Mill and secure the cover by tightening the clamps.
4. Position a sample collection bottle under the cyclone body.
5. Plug the power and turn on the mill. Make sure the noise is normal when the mill is on.
6. Slowly add the samples from the top and stop adding samples when the collection bottle is 2/3 full.
7. Turn off the mill and unplug the power.
8. Collect the fine powders.
9. Open the mill and clean it using brushes and vacuum.



### iii. Cut mill

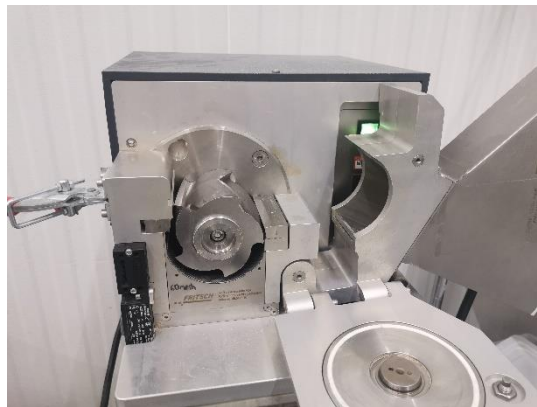
Preliminary tests show that the cut mill is a good device to mill the pulses as a whole into fine powders. Milling yield is about 90%. The procedure describes the steps of milling using the cut mill.

#### Apparatus:

1. Cut mill with #60 mesh sieve
2. Balance
3. Brushes
4. Vacuum
5. N95 mask
6. Ear plug or ear muff

#### Procedure:

1. Weigh out 220 g seeds (without foreign matter).
2. Switch the mill to AUTO mode. Open the milling chamber once the green light is on.
3. Make sure the chamber is cleaned and the #60 mesh screen is used. Close the chamber.



4. Turn on the mill. Make sure the noise is normal when the mill is on.
5. Put the samples from the filling chamber. Only put a full hand of seeds every time, and wait for about 30 seconds. Listen the voice to determine whether milling is finished. Do not put all seeds at once; otherwise, extensive heat will be generated.
6. Wait for one minute when the last samples are injected. It takes about 6 minutes for milling 170 g of lentils and about 9 minutes to mill one chickpea sample.
7. Stop the mill and collect the sample.
8. Weigh the resulted flours and record the value.
9. Open all chambers and clean out the remained samples using brushes and vacuum.

#### iv. Stone mill

Preliminary tests show that stone mill is a good device to separate the hull and the cotyledon. The procedure describes the steps for dehulling and milling.

##### Apparatus:

1. Stone mill
2. Balance
3. Brushes
4. Vacuum
5. Powder sifter machine (#35 and 60 meshes)

##### Procedure:

1. Weigh out 220 g seeds (without foreign matter).
2. Make sure the mill is cleaned before milling.



3. Turn on the mill and make the sure it is functioning properly by listening the noise.
4. Slowly add the samples from the top and wait till all samples are milled.
5. Stop the mill and collect the flours.
6. Put the samples the sifter machine and start sifting at level 6 for 15 min.
7. Open the mill and clean out all the remained powders using brushes and vacuum.
8. Check the samples at each screen once sifting is done.

## I. Colour of pulse flour

This procedure describes the steps to measure the absolute colour of the pulse flour ( $L^*$ : white (100) to black (0),  $a^*$ : red (+) to green (-),  $b^*$ : yellow (+) to blue (-)). Values indicate the visual quality of the flour and the processing effect on colour. Lightness of flour increases as the fineness increases.

### Apparatus:

1. Konica Minolta CR-400 Chroma meter
2. Petri dish
3. Spatula
4. Cleaning foam brush

### Procedure:

1. Connect the measuring head and data processor with the RS-232C cable.
2. Lock the measuring head with the stand.
3. Switch the power ON for the measuring head then data processor. Wait till **DP MODE** is displayed in the measuring head.
4. Press the **Display** on the data processor till  $L^*, a^*, b^*$  are displayed.



5. White calibration: take off the protective cap from the measuring head and place the calibration plate vertically. Press **calibration** on the data processor. The screen should appear **Y=93.5, x=0.3114, y=0.3190**, or use the numeric pad to set the correct values. Press **measure enter**, and the screen will show **NOW CALIBRATING**.



6. Absolute measurement: fill up the petri dish with flours needed to be measured. Put on the petri dish holder, petri dish, and the lid to the measuring head. Press **measure enter** on the data processor. Three measurements are made each time, and the average is displayed on the screen. **Always make sure there is no powder dropped into the measuring head.**

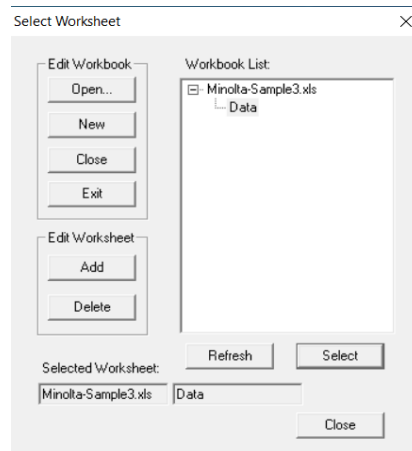


7. Turn off both devices when measurement is done.
8. Put every piece back to the case.



#### **Data download:**

1. Connect data processor to PC with USB-Serial Converter Cable. Switch the power ON and press **Enter** to select **REMOTE MODE**.
2. Click on software “CR-400 Utility” from the PC. From the window, choose **Sensor head** in **Connect to** frame. Click **Port Setting**, choose **COM7** in Port and **12600** in Baudrate; then click OK. Click **Connect** to connect the PC and measuring head.
3. Choose **Excel Spreadsheet** in Target to redirect data. In the **Select Worksheet** frame, click Open and choose **Minolta-Sample 1**. In the “Workbook List”, click **Data** and click **Select**, then **Close**.



4. Click **Instrument > Upload**. In the “Upload Data” frame, click **Upload**, and all the data stored in the measuring head is now displayed on the excel sheet.

**Note:**

1. Clean the petri dish properly between each sample. **Always make sure there is no powder dropped into the measuring head.**
2. The measuring head can store up to 1000 data. The device will delete data automatically after exceeding the limit.
3. Make sure the data are deleted in the device after uploading to the Excel sheet.



## J. Hausner ratio

Hausner ratio measures the ratio of tapped density to loose bulk density, indicating the flow-ability and the compressibility of the flour after milling. Hausner ratio is an important parameter in food products handling, packaging, storage, processing and distribution. It is useful in the specification of products derived from size reduction or drying processes. Usually, the lower the flow-ability a flour, the more compressible it becomes<sup>5</sup>.

### Apparatus:

1. 25 mL graduated cylinder
2. Spatula
3. Funnel

### Procedure:

1. Place a 25mL cylinder on a scale. Let 10.0 g of flour loosely fall in the cylinder by using a funnel.
2. Record the exact volume (i.e., start volume) of the flour till 2 decimals.
3. Lightly tap the cylinder 40 times.
4. Increase the strength, with dropping motion, tap for 60 times then stop to check.
5. Tap like Step 3 for 30 times more. Record the reading (i.e., end volume) to 2 decimal place.
6. Duplicate the measure for each sample.

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<sup>5</sup> Buanz, A. (2021). Powder characterization. In *Remington* (pp. 295-305). Academic Press.  
<https://doi.org/10.1016/B978-0-12-820007-0.00016-7>

Amankwah, N. Y. A., Agbenorhevi, J. K., & Rockson, M. A. (2022). Physicochemical and functional properties of wheat-rain tree (*Samanea saman*) pod composite flours. *International Journal of Food Properties*, 25(1), 1317-1327. <https://doi.org/10.1080/10942912.2022.2077367>

Aulton, M. E., & Taylor, K. M. G. (2013). *Powder flow* (pp. 189-200). Edinburgh, Scotland: Churchill Livingstone (Elsevier).

Maninder, K., Sandhu, K. S., & Singh, N. (2007). Comparative study of the functional, thermal and pasting properties of flours from different field pea (*Pisum sativum* L.) and pigeon pea (*Cajanus cajan* L.) cultivars. *Food chemistry*, 104(1), 259-267.  
<https://doi.org/10.1016/j.foodchem.2006.11.037>

Ogunsina, B. S., Radha, C., & Govardhan Singh, R. S. (2010). *Physicochemical and functional properties of full-fat and defatted Moringa oleifera kernel flour*. *International Journal of Food Science & Technology*, 45(11), 2433–2439. <https://doi.org/10.1111/j.1365-2621.2010.02423.x>

**Calculation:**

$$\text{Loose bulk density} = \frac{m (g)}{\text{start volume (ml)}}$$

$$\text{Tapped density} = \frac{m (g)}{\text{end volume (ml)}}$$

$$\text{Hausner ratio} = \frac{\text{Tapped density}}{\text{loose poured density}} = \frac{\text{start volume (mL)}}{\text{end volume (mL)}}$$

$$\text{Compressibility} = \frac{\text{Tapped density} - \text{Loose bulk density}}{\text{Tapped density}} \times 100\%$$

**Note:**

1. When pouring the flour into the cylinder, use the wall of the funnel to avoid direct dropping into the cylinder to prevent pre-packing the flour.

**Table 1j.** Relationship between powder flow-ability, percentage compressibility and Hausner ratio.

Compressibility index (%) (Carr's index)	Type of flow	Hausner ratio
1-10	Excellent	1.00-1.11
11-15	Good	1.12-1.18
16-20	Fair	1.19-1.25
21-25	Passable	1.26-1.34
26-31	Poor	1.35-1.45
32-37	Very poor	1.46-1.59
>37	Very, very poor	>1.59

## K. Particle size

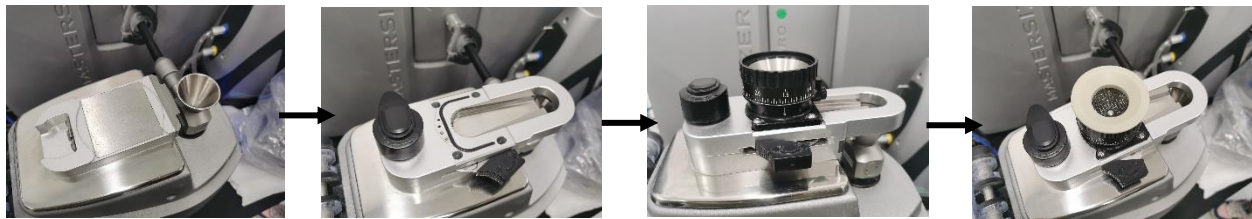
This protocol describes the procedure to measure the average particle size and droplet size distribution for dry powders.

### Apparatus:

1. Mastersizer 3000 with dry sample cell
2. Spatula
3. Cleaning brushes

### Procedure:

1. Push the power switch to turn on the Mastersizer 3000, and wait for 30 min for the instrument to warm up. Turn on the air to 5.5 bar. Air will only be turned on for dry samples.
2. Set up all three pieces of apparatus for loading dry flours with the order showing below.



3. The gap should be set based on the flour conditions. Setting the gap to 3 is a good starting point for lentil, pea, faba bean, dry bean, and chickpea (black desi and desi types) flours (gap of 3.5 for chickpea-kabuli flours).
4. Close the sample cell lid, and make sure it is sealed.
5. From the PC, click on software **Mastersizer 3000**.
6. Cleaning: from Menu, choose **Tool > Accessories > Clean > Extensive**. Click **Clean** to start cleaning. You will hear three different noises as cleaning proceeds.
7. Go to **Home > Manual measurement**. In the setting, name the sample and make sure the setting is in default.

Particle size: non spherical

Material:

- silica flour
- refractive index: 1.56
- absorption index: 0.01
- density: 1

8. Click **Initialise instrument**, make sure the highest point of Energy do not exceed 250. Otherwise, a second cleaning is required. Click **stop** when the condition is ok.
9. Open the sample cell lid and put a full cup of samples. Close the lid.
10. In Menu, click **air flow** to start the air.

11. Click **feed rate** and start at about 28% for lentil, pea, faba bean, dry bean, and chickpea (black desi and desi types) flours and 33% for chickpea-kabuli flours. Make sure the obscuration stays in green by changing the feed rate.
12. Start **measurement**. You should monitor the obscuration all the time and change the feed rate if it is not in green.
13. Five measurements are preformed for each sample. The results will show in the screen, and the SD for all parameters should be less than 5%. Otherwise, a repeat is needed. Click **Stop** when the measurement is finished.
14. Select all five results and click **Data quality**. Make sure all the results are in good quality. Otherwise, a repeat is needed.
15. Cleaning: brush out all the remained samples from the dry sample cell. In Menu, click **Clean** again (see step 1) and check **initialise instrument** (see step 8).
16. Now it is ready for a new measurement.
17. Result saving: click “save as” and name the file based on data.
18. By the need of the day, take off the loading cell and rinse them properly.
19. Turn off the instrument and the air control.

**Result upload:**

1. Open the saved file and transfer all the average value and SD to an excel sheet.
2. For the size distribution, in “exported” frame, click **export content** then **copy as image** and paste the image to Word.

## L. Ash content

This method determines the ash (total mineral) content of pulse flour. The method is modified from AACC 08-01.01<sup>6</sup>.

### Apparatus:

1. Muffle Furnace
2. Analytical balance
3. Crucibles
4. Desiccator
5. Tong
6. Heat-resistant gloves

- **Daily Loads: 3**
- **# Samples per load: 9**



### Procedure:

1. Label a number at the bottom of the crucibles using pencils.
2. Place a crucible on the balance and tare.
3. Weigh  $0.32 \pm 0.02$  g samples in the crucibles ( $0.22 \pm 0.02$  g for chickpea flours) and record the sample weight.
4. Prepare 9 samples for the 1<sup>st</sup> load.
5. Place all the crucibles into the desiccator.
6. Place the crucibles in the muffle furnace.
  - a. The crucibles CANNOT touch each other;
  - b. The crucibles CANNOT touch the inside wall of the furnace.
7. Close the door, turn on the furnace, and:
  - a. Set the temperature to **560 °C** for **15 min** and turn on the fume hood.
  - b. After 15 minutes, set the timer to **2 h 25 min**.
  - c. Turn off the fume hood after 45 minutes.
8. Weigh samples for the 2<sup>nd</sup> and 3<sup>rd</sup> loads. Put all the samples in the desiccator.

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<sup>6</sup> AACC (1999). American Association of Cereal Chemists International. Approved methods of analysis (11th ed.). The Saint Pauls Association: Saint Paul, MN.



9. When the 1<sup>st</sup> load is done:
  - a. Turn off the furnace.
  - b. Take the 2<sup>nd</sup> load of samples out of the desiccator.
  - c. Wear heat-resistant gloves and use the tong to take the 1<sup>st</sup> load of samples out of the furnace and place them in the desiccator.
  - d. Wear heat-resistant gloves and use the tong to put 2<sup>nd</sup> load of samples in the muffle furnace.
  - e. Close the furnace door.
  
10. Weigh the 1<sup>st</sup> samples (and 3<sup>rd</sup> load from the previous day; see step 14) after **1 h** in the desiccator.
  - a. Use a **tweezer** to handle the crucible.
  - b. Record the total weight (crucible + sample).
  - c. Discard the ash by using a tweezer. Wipe off any remaining ash.
  - d. Record the weight of the empty crucible in the crucible weight section.
  
11. Repeat step 7, step 9, and step 10 to finish the 2<sup>nd</sup> and 3<sup>rd</sup> loads.
  
12. Turn off the furnace at the end of Round 3. Prepare a next-day 1<sup>st</sup> load of samples and put them in the furnace. The remaining heat will burn off a part of the samples.
  
13. Sample crucibles of the 3<sup>rd</sup> load will be left in the desiccator overnight.
14. Weigh the 3<sup>rd</sup> load of samples and the next-day 1<sup>st</sup> load **together** on the next day.

**Calculation:**

$$\text{Ash Content (\%)} = \frac{\text{Weight of the crucible + sample after ashing (g)} - \text{weight of crucible (g)}}{\text{weight of the sample before ashing (g)}} \times 100\%$$

**Note:**

1. Furnace life is reduced by repeated heating and cooling.
2. Keep the furnace above 350 °C between runs.
3. Brown coloration on the insulation material does not affect the performance/functionality of the furnace.

## M. Moisture content

This method determines the moisture content of pulse flour, modifying from AOAC 925.10<sup>7</sup>.

### Apparatus:

1. Analytical balance
2. Convection oven
3. Aluminum moisture pan
4. Aluminum tray
5. Desiccator
6. Tweezers/ gloves

### Procedure:

1. Label each moisture pan (36 pans in a run).
2. Place all moisture pans on a tray, and place the tray on the **top layer** of the oven at 130 °C for 30 min.
3. Take the tray from the oven and place the tray in the desiccator for 15 min.
4. Weigh a moisture pan and record the weight to 4 decimals. Do not touch the moisture pans with your hands to avoid transferring moisture/lipid (use a tweezer or gloves).
5. Weigh 1.2 ± 0.02 grams of samples in the moisture pan. Record the sample weight.
6. Place the sample on a **new tray**.
7. Put the tray with all samples in the oven (**middle layer**) at 130°C for 2 h 10 min. Do not put the samples on the bottom layer, as it will burn the samples. Check every 30-min to make sure the temperature maintains at 130 °C.
8. Take the samples from the oven and put them in the desiccator for 1 hour.
9. Weigh the sample + pan.
10. Duplicated measurements are performed for each flour.

### Calculation:

$$\text{Moisture content (\%)} = \frac{\text{moisture pan weight (g)} + \text{sample weight before drying (g)} - \text{weight of pan + sample after drying (g)}}{\text{weight of sample before drying (g)}} \times 100\%$$

### Notes:

1. Make sure the desiccator is properly closed and desiccant is dry (blue = good; pink/purple = wet).
2. Do not put the desiccator on the weighting bench.

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<sup>7</sup> AOAC. (2005). Official method of analysis (27th ed.). Virginia: Association of Official Analytical Chemists Inc.

## N. Protein content

Protein content is determined through the combustion method (AOAC 990.03). Nitrogen is detected through a thermal conductivity detector and converted to protein content with a protein factor of 6.25.

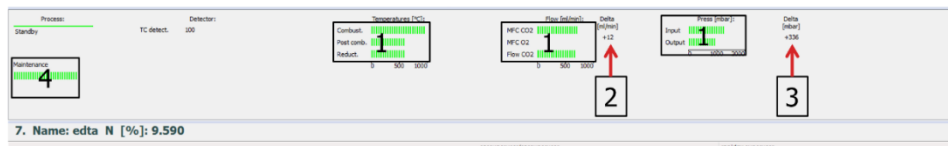


### Apparatus:

1. Protein Analyzer (Elementar, German)
2. Analytical balance
3. Weighing apparatus
4. Heat resistance gloves, tong, pliers, wrench, and ladder

### System Preparation:

1. Record your name and start time in the Log Book.
2. **If the instrument is off** from previous day, turn on the analyzer by pressing the green power button on the lower right side. Open the software and wait for the connection to be established. Turn on the furnace through **System > Furnace**. Wait till the furnace temperature reach 950 °C and give another half an hour for stabilization.
3. **If the analyzer is on** from the previous day, wake up the system from sleep mode using the menu **System > Wake up**.
4. Turn on both gas tanks by turning a quarter ( $\frac{1}{4}$ ) of the knob directly connected to each gas tank. The direction of opening and closing is indicated on top of the knob.
5. The intake pressure of the O<sub>2</sub> is 2.5 bar, and CO<sub>2</sub> is 1.4 bar. The pressure generally remains stable, and usually you don't need to make any adjustment (or adjust it slightly). However, you should monitor the values throughout the day. Report to your supervisor if a large discrepancy occurs.
6. When the above steps are ready, go through the standby checklist.
  - a. Make sure the furnace temperatures, flows, and pressures all green.
  - b. Flow delta < 15 ml/min.
  - c. Pressure delta < 400- 500 mbar. (Up to 500 mbar is still okay, but the furnace tubes should be replaced soon).
  - d. Maintenance counter bar green.

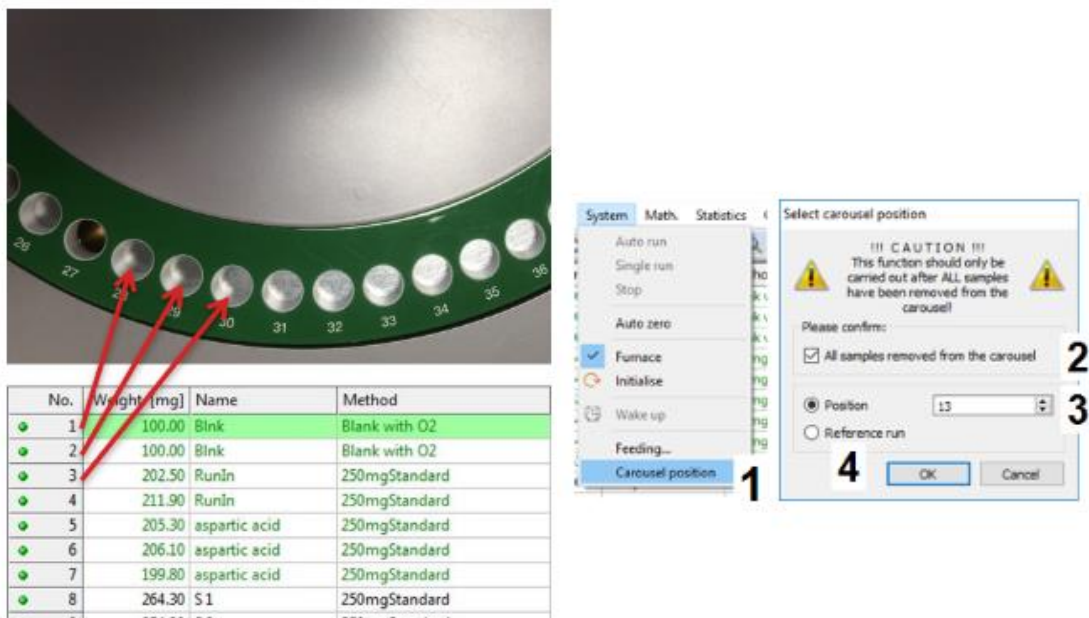


### Sample Preparation:

7. Everyday you should create a new file from the menu **New**. Save the file in G drive and name it with the following format: YYYYMMDD-seed type-location-protein (e.g. 20230606-pea-Swan River-protein). Save the file multiple times during the day to ensure your progress is consistently saved.
8. There are three types of runs for preparing the analyzer: **blank**, **run-in**, and **daily factor**. Samples prepared should follow the below list: 2 run in of blank, 3 blank, 2 run in of aspartic acid, and at least 3 aspartic acid.
  - a. **2 run in of blank:** the first N area is usually high, and the second value normally is below 150.
  - b. **3 blank:** all N area < 150. The three values will be used as blank value in the calculation. You should make sure all blanks are at the range before you proceed further.
  - c. **2 run in of aspartic acid:** for analyzer to run, not taking in the daily factor calculation.
  - d. **Aspartic acid:** N factors should be between 0.900 and 1.100. The largest difference between any two factors should be less than 0.010 (ideally < 0.005). Or you must perform more aspartic acid until you have 3 N factor values with differences smaller than 0.01.
  - e. It is recommended to put a **stop tag** at the end of aspartic acid. You should keep an eye of each O<sub>2</sub> curve (**square-shaped oxygen flow**).
  - f. Routine blank, run-in, and daily factor should be performed again after replacing part.

No.	Weight [mg]	Name	Method	N Area	N [%]	N Factor	N Blank	Protein [%]	Protein Factor
2 Run In	100.00	RunIn	Blank with O2	95	0.031	1.0000	0	0.196	6.2500
3	100.00	RunIn	Blank with O2	72	0.014	1.0000	0	0.090	6.2500
3 Blank	100.00	Blnk	Blank with O2	52	0.000	1.0000	52	0.000	6.2500
5	100.00	Blnk	Blank with O2	66	0.000	1.0000	66	0.000	6.2500
7	100.00	Blnk	Blank with O2	68	0.000	1.0000	68	0.000	6.2500
2 Run In	250.70	RunIn	250mgStandard	37 587	10.597	1.0000	62	66.234	6.2500
10	250.10	RunIn	250mgStandard	37 415	10.574	1.0000	62	66.089	6.2500
3 Aspartic acid	250.20	aspartic acid	250mgStandard	38 005	10.520	0.9798	62	65.750	6.2500
8	251.20	aspartic acid	250mgStandard	37 833	10.520	0.9882	62	65.750	6.2500
10	249.70	aspartic acid	250mgStandard	37 885	10.520	0.9809	62	65.750	6.2500
11	125.50	308	250mgStandard	5 809	3.200	0.9830	62	20.003	6.2500
12	125.10	108	250mgStandard	6 484	3.586	0.9830	62	22.412	6.2500
13	125.20	108	250mgStandard	6 470	3.575	0.9830	62	22.346	6.2500
14	125.30	205	250mgStandard	6 222	3.435	0.9830	62	21.466	6.2500
15	124.90	205	250mgStandard	6 163	3.413	0.9830	62	21.328	6.2500
16	126.00	303	250mgStandard	6 194	3.400	0.9830	62	21.250	6.2500

9. Always check whether the balance is level. Adjust the leveling feet until the bubble is exactly in the center of the level indicator. Always keep the balance clean and clean it immediately after a spill.
10. Using manual pressing tool, lightly press the tin foil into bowl shape (**50 x 50 mm** for run-in and aspartic acid; **35 x 35 mm** for pulse flour), place the pressed tin foil onto the balance, close the door, and tare the balance.
11. Add the sample to the container (0.249-0.251 g for run-in and aspartic acid; 0.1235-0.1265g for pulse flour).
12. Enclose the sample using the manual pressing tool.

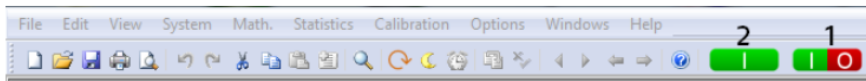


13. Place the enclosed sample on the balance, close the balance doors, and record the sample mass to the software by clicking **Print**. Make sure the current weight (yellow line) is correct, or right click the desire line and select **Current weight**.
14. Place the sample on the carousel. Make sure you match the carousel position with the software (Ball valve position + 1).
15. If you would like the measurement to **start at position 1**, adjust the carousel position through **System > Carousel position**. The position will be at #60 so to start at #1 (with 2 run-in, 3 blank, actual placement begins at #5). Make sure you remove all the samples from the carousel before adjustment.

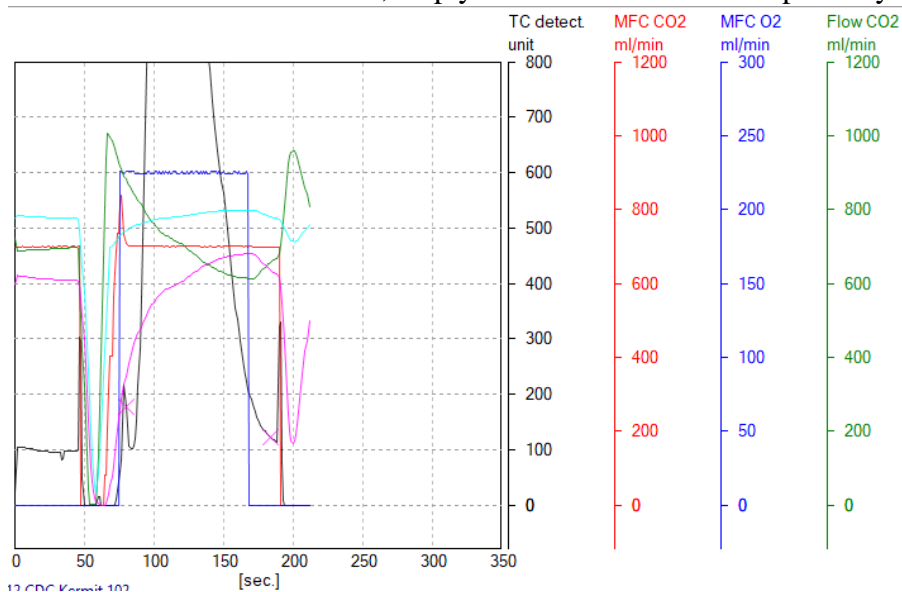


**Sample Measurement:**

16. Now you are able to start your sample measurement. Click **System > Single run (1)** or **System > Auto run (2)**.



17. Always check your results, and gas flows on graphs as the measurement is running.
  - a. Always check for square-shaped oxygen flow.
  - b. CO<sub>2</sub> and O<sub>2</sub> gas flow: they should look similar to the picture below; any large fluctuation of these two gases can cause the analysis to be inaccurate due to incomplete combustion.
  - c. If a block of O<sub>2</sub> is observed, stop your measurement and report to your supervisor.



18. Based on setting, the analysis will stop at the last sample (or sample with a stop tag) when you use auto run, and the system will turn to sleep mode.
19. The system will then start depressurizing. You are able to turn off both gas tanks after that.
20. Please **DO NOT** close the software or shut down the analyzer. **DO NOT** turn off the computer.

**Save and Export the file:**

21. Save your file at the beginning of the day and save it multiple times during measurement.
22. Export the file and Excel sheet to G drive by the end of the day.
  - a. Click **File > Export/ Import > Export** to export the file as SQLITE file.
  - b. Click **File > Export/ Import > Export sheet data (for excel)** to save an Excel data sheet.

**Log Book Rand Cleaning:**

23. Record information in the Log Book: time, # of sample, condition, and write down any maintenance performed or situation.
24. Clean and vacuum your work station by the end of the day.

**Shut Down Instrument:**

25. When a measurement break is longer than a week. Talk to your supervisor before shutting down.
26. Shut off gas supply (**step 19**).
27. Switch off the furnace in the operating software through **System > Furnace**.
28. Wait 2-3 hours until the instrument has cooled down to < 55 °C.
29. Quit the software through **File > Exit**.
30. Switch off the analyser and its peripherals.

**Safety and Maintenance of Analyzer:**

1. You **must** perform any of the maintenance activity when there are two people around.
2. You **must** use a ladder, wear heat resistance gloves, and use proper tools when performing maintenance activity.
3. Report all troubleshooting activities to your supervisor and record in the Log book.
4. Review the proper steps and video for each maintenance/troubleshooting activity from **Elementar Academy** previous to any implement.
5. Maintenance work that requires opening gas lines should only be performed when the analyzer is not pressurized.
  - a. To de-pressurize the analyzer for maintenance, use the menu: **Options > Maintenance > Replace parts**
6. The current maintenance status is displayed in the lower left corner of the status view.
  - a. Green: <80% of interval
  - b. Orange: ≥80% of interval
  - c. Red: ≥100% of interval
7. Check the status of each event by clicking on: **Options > Maintenance > Intervals**
8. It is usually not necessary to shut down the analyzer or to cool down the furnace before performing maintenance actions, such as:
  - a. Performing a leak check
  - b. Emptying or exchanging the ash crucible
  - c. Exchanging combustion and drying tubes
  - d. **Except** for removing the ball valve. This requires a **cool furnace**.
9. Spare reaction or drying tubes can be filled prior to replacing parts, thus reducing the downtime of the analyzer.

**Important:**

4. Furnace life is reduced by repeated heating and cooling.
5. Keep the furnace at 950°C. Do not shut it off for weekly use.
6. DO NOT change the furnace temperature.
7. DO NOT change the standard method. If a new method is needed, report to your supervisor.
8. DO NOT change the sample size. Report to your supervisor if a sample size needs to be adjusted.

**Maintenance Action List:**

Note that the default interval might be changed according to the actual conditions.

<i>MAINTENANCE ACTION</i>	<i>DEFAULT INTERVAL</i>
<i>Clean the nickel flap</i>	<i>150 samples</i>
<i>Empty or exchange the ash crucible</i>	<i>150 samples</i>
<i>Refill the eas regainer®</i>	<i>500 samples*</i>
<i>Replace the large drying tube filling</i>	<i>300 samples</i>
<i>Replace the small drying tube filling</i>	<i>1000 samples</i>
<i>Replace the combustion tube filling</i>	<i>1000 samples</i>
<i>Replace the post-combustion tube filling</i>	<i>2000 samples</i>
<i>Replace the eas reductor®</i>	<i>2000 samples</i>
<i>Clean the ball valve</i>	<i>2000 samples</i>
<i>Clean the quartz bridge</i>	<i>Inspect daily</i>
<i>Clean the water condenser</i>	<i>Inspect monthly</i>
<i>Replace the combustion tube</i>	<i>5 fillings</i>
<i>Replace the post-combustion tube</i>	<i>20 fillings</i>
<i>Replace the ash crucible</i>	<i>2000 samples</i>
<i>Replace the oxygen lance</i>	<i>2000 samples</i>
<i>Replace o-rings and seals</i>	<i>Anually</i>
<i>Clean and adjust the autosampler</i>	<i>Anually</i>
<i>Clean analyzer and check for worn-out parts</i>	<i>Anually</i>
<i>Recalibrate the TCD</i>	<i>Anually or less</i>

### **3. INSTRUMENT MAINTENANCE**

This section describes the instrument maintenance, safety instruction, and other important notes to each instrument.

#### **A. Balance**

##### **Capacity:**

The analytical balance and the top loading balance have the maximum capacity of 120g and 5000g, respectively. Do not weight anything that passes the limit.

##### **Transporting:**

1. Always proceed focused and with care.
2. Hold the balance with both hands. Do not lift it by the glass draft shield.
3. The balance should be placed to avoid vibrations and temperature fluctuations. Do not place the balance and the powder sifter machine at the same bench. Do not place the balance near the muffle furnace or oven.

##### **Leveling:**

1. Before using the balance (both analytical and top loading balances), make sure the air bubble is within the circle of the level indicator. If not, use the level feet on both sides to adjust the indicator position.

##### **Cleaning:**

1. Improper cleaning can damage the load cell or other essential parts. Do not use any cleaning agents. Do not spray or pour any liquids to the balance.
2. Use a moistened lint-free cloth or a tissue.
3. Remove any dirt or dust around the balance.
4. Brush out any dust or samples dropped in the balance after every use.
5. Check the level status after cleaning

##### **Calibration:**

1. The analytical balance comes with internal calibration.
2. Weigh an item with known weight daily is recommended.

## **B. Muffle furnace**

### **Capacity:**

1. The maximum temperature for the furnace is 1100°C. Do not run the furnace above 1000°C.

### **Cleaning:**

#### *Daily:*

1. Remove any dust in the chamber before any use.
2. Keep the furnace clean before and after use.

#### *Monthly:*

3. Run the empty furnace at 850 °C for 8 hours to burn off any contamination on the insulation and elements.

### **Important:**

1. Furnace life is reduced by repeated heating and cooling.
2. Keep the furnace above 350 °C between runs.
3. Brown coloration on the insulation material does not affect the performance/functionality of the furnace.

### **Thermocouple:**

1. Replacing the thermocouple periodically (once every six months) to ensure temperature accuracy is recommended.



## **C. Colourimeter**

### **Operation environment:**

1. Do not use the instrument near source of heat, flammable gas, dust, or magnetic field.
2. Use the instrument at ambient temperature between 0 and 40°C.
3. Do not subject the instrument to strong impact or vibration. Always attach the protective cap when not in use.
4. Always lock the measuring head in the stand while in use.

### **Storage:**

1. Always put the instrument back to the case while not in use.

### **Cleaning:**

1. Always keep the instrument clean.
2. Wipe off any dust with a soft, clean dry cloth.
3. Do not use any solvents for cleaning.
4. White calibration plate can be cleaned by a soft, clean dry cloth. Use lens cleaner and dry with cloth when the dirt is difficult to remove.

### **Calibration:**

1. Calibrate the instrument prior to every use with the white calibration plate.
2. Do not allow calibration plate to get scratched or stained.

## D. Protein analyzer

### Safety:

1. You must perform any of the maintenance activity when there are two people around.
2. You must use a ladder, wear heat resistance gloves, and use proper tools when performing maintenance activity.

### Maintenance of Analyzer:

1. Report all troubleshooting activities to your supervisor and record in the Log book.
2. Review the proper steps and video for each maintenance/troubleshooting activity from **Elementar Academy** previous to any implement.
3. Maintenance work that requires opening gas lines should only be performed when the analyzer is not pressurized.
  - a. To de-pressurize the analyzer for maintenance, use the menu: **Options > Maintenance > Replace parts**
4. The current maintenance status is displayed in the lower left corner of the status view.
  - a. Green: <80% of interval
  - b. Orange: ≥80% of interval
  - c. Red: ≥100% of interval
5. Check the status of each event by clicking on the bar or using the menu: **Options > Maintenance > Intervals**
6. It is usually not necessary to shut down the analyzer or to cool down the furnace before performing maintenance actions, such as:
  - a. Performing a leak check
  - b. Emptying or exchanging the ash crucible
  - c. Exchanging combustion and drying tubes
  - d. **Except** for removing the ball valve. This requires a cool furnace.
7. Spare reaction or drying tubes can be filled prior to replacing parts, thus reducing the downtime of the analyzer.

### Important:

1. Furnace life is reduced by repeated heating and cooling.
2. Keep the furnace at 950°C. Do not shut it off for weekly use.
3. DO NOT change the furnace temperature.
4. DO NOT change the standard method. If a new method is needed, report to your supervisor.
5. DO NOT change the sample size. Report to your supervisor if a sample size needs to be adjusted.